**Acinetobacter Minimal Succinate Media Recipe (courtesy of the Averhoff lab, with modifications for available reagents)**

To make 1L of standard Acinetobacter Minimal Succinate Media (ABMS), combine the following:

* 880mL autoclaved diH2O, room temperature
* 20mL of 1M Sodium Succinate (sterile, room temperature)
* 50mL of 20X Mineral Solution (sterile, room temperature)
* 50mL of 20X Phosphate Buffer (sterile, room temperature)

To make 1L of AMBS plates:

* Add 16g/L agar to dH2O before autoclaving in a 2L flask.
* Warm components at 55C in water bath (to reduce temperature difference when added – optional step)
* Once water and agar are cool enough to touch the flask, add component solutions.

Note: 50C

Component Recipes

**SL9 (trace minerals)**

* 800ml diH2O
* Nitrotriacetic acid 12.8g
* FeSO4•7H2O 2g (note – dissolve first in 2N HCl)
* CoCl2 (anhydrous) 104mg
* MnCl2•4H2O 122mg
* ZnCl2 70mg
* NaMoO4•2H2O 36mg
* NiCl2 13mg

And 1ml of 10ml 10X solution

* H3BO3 60mg
* CuCl2•2H2O 20mg

Bring to 1000mL and pH 6.5. Note – this requires a lot of NaOH, so leave room when bringing up to volume.

**Mineral Solution (20x)**

* NH4Cl 10g
* MgSO4•7H2O 5.9g
* KNO3 1g
* (NH4)6Mo7O24•4H2O 20mg
* CaCl2 Solution 931.5µl
* 10ml of SL9

Brought up volume to 500ml in graduated cylinder with deionized water.

**Phosphate Buffer (20x)**

* KH2PO4  68g
* Na2HPO4•H2O 132.5g
* 400ml deionized water, pH to 6.8
* Volume brought up to 500ml